

## *Short Communication*

# First and second derivative spectrophotometric assay of mixtures of cefuroxime and cephalexin

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### **Introduction**

Derivative spectrophotometry offers a convenient solution to a number of well defined analytical problems, such as resolution of multicomponent systems, elimination of interference from sample turbidity and matrix background, and enhancement of spectral details [1–3].

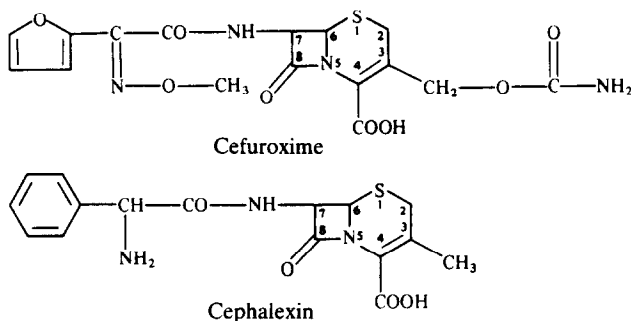
Fell [4] discussed the use of the derivative technique for resolution enhancement in HPLC. The technique involves differentiating the output of the detector, which may be based on UV absorbance, fluorescence, etc., with respect to time to give the second or fourth derivative.

One of the classical analytical problems in the field of UV–visible spectrophotometry is the resolution of a number of components mixture with overlapping bands. A number of papers have appeared describing applications of derivative spectrophotometry in the analysis of mixtures of metals. For example, satisfactory derivative methods for the simultaneous determination of mixtures of bismuth(III)–copper(II) [5], iron(III)–copper(II) [6] and ruthenium(III)–palladium(II) [7], using 2-thiobarbituric acid as the colorimetric reagent and osmium(VIII)–palladium(II) [8], using allyl thiourea as the reagent. First derivative spectrophotometry has recently been employed for the determination of mixtures of iron(III)–bismuth(II) using EDTA as the reagent [9].

Derivative spectrophotometry has also proved to be useful in the assay of many drugs. Fell has discussed the relative ease of second derivative UV–visible spectrophotometry for the analysis of pharmaceutical dosage forms [10] and the rationale for developing bioanalytical procedures based on higher derivative methods [11]. First and second derivative spectrophotometry recently have been employed for the determination of certain drugs, e.g. acepifylline and phenobarbitone [12], some cephalosporins [13] and pirbuterol and butorphanol [14].

For the determination of substances of clinical interest [15–19], first and second derivative spectrophotometry has been employed for the simultaneous determination of cefuroxime and cephalixin, which are cephalosporins with closely overlapping absorption spectra. The chemical structures of these antibiotics are shown in Table 1. It is the purpose of this work to demonstrate the ease with which the method overcomes the problem due to overlapping spectral bands and to prove that it is a very simple and useful procedure for determining the two cephalosporins in a mixture without tedious and time-consuming separation procedures.

**Table 1**  
Chemical structures of cefuroxime and cephalixin



## Materials and Methods

### Reagents

Solutions of cefuroxime sodium and cephalixin sodium ( $0.2 \text{ mg ml}^{-1}$ ) in water were freshly prepared.

### Apparatus

A Perkin–Elmer 555 double-beam UV–visible spectrophotometer was used.

The optimisation of derivative spectra requires the selection of a suitable slit width, response time and scan speed of the monochromator. A spectral bandwidth of  $1 \text{ nm}$ , a response time of  $4 \text{ s}$  and a scan speed of  $120 \text{ nm min}^{-1}$  were found to be optimum for both 1st and 2nd derivative spectra. The recorder scale expansion was also optimised to facilitate readings on the recorder tracing.

### Procedure

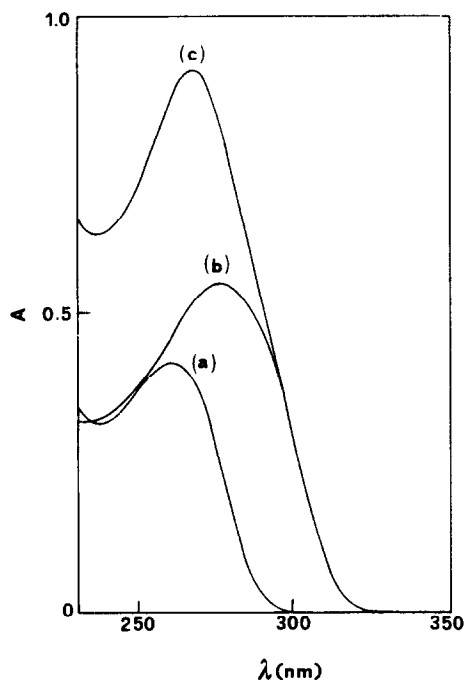
A few  $\mu\text{l}$  of the cephalosporins solutions were mixed in a 5-ml calibrated flask and diluted to volume with distilled water. The spectrophotometric measurements were performed against water in 1-cm quartz cells.

## Results and Discussion

### Spectrophotometric measurements

Figure 1 shows the absorption spectra of: (a) cephalixin sodium ( $20 \mu\text{g ml}^{-1}$ ) with a maximum at  $261 \text{ nm}$ ; (b) cefuroxime sodium ( $15 \mu\text{g ml}^{-1}$ ) with a maximum at  $275 \text{ nm}$  and; (c) a mixture of cephalixin and cefuroxime ( $20 \mu\text{g}$  and  $15 \mu\text{g ml}^{-1}$ , respectively) with a maximum located at  $265 \text{ nm}$ , i.e. between the absorption maxima of the two

**Figure 1**  
Zero-order spectra of: (a) cephalixin sodium; (b) cefuroxime sodium; and (c) cephalixin sodium plus cefuroxime Na. cephalixin sodium,  $20 \mu\text{g ml}^{-1}$ ; cefuroxime sodium,  $15 \mu\text{g ml}^{-1}$ ; reference, water.



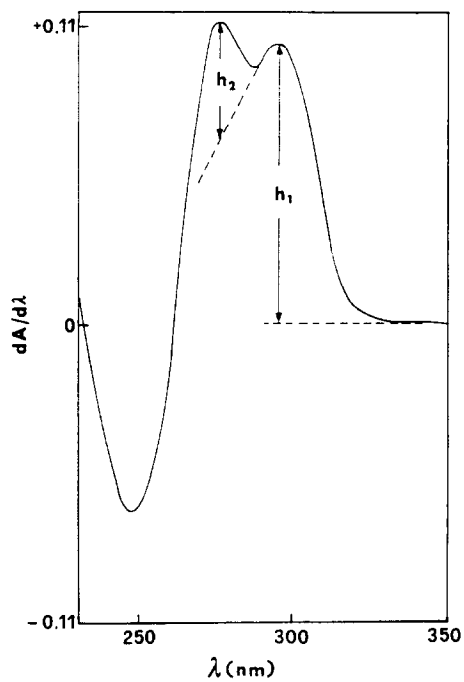
components. Thus the overlap of the spectra of cefuroxime and cephalixin result in the absence in the zero-order spectrum of the mixture of any spectral feature (e.g. shoulder) that can be used to assay the two substances. Moreover, the traditional Vierordt's method used for assaying binary mixtures which involves the use of two simultaneous equations, and its variant, the modified Vierordt's method, are time-consuming and tedious, and lack precision when the  $\lambda_{\text{max}}$  are close together [10, 12–14].

These difficulties were overcome by the use of derivative spectrophotometry. Figure 2 is the first derivative spectrum of the zero-order spectrum of the mixture shown in Fig. 1. Figure 3 shows a typical second derivative spectrum of another mixture ( $30 \mu\text{g ml}^{-1}$  of cefuroxime and  $20 \mu\text{g ml}^{-1}$  of cephalixin). In contrast to the zero-order spectrum, the first derivative spectrum gives separate peaks at 296 and 276 nm. The second derivative spectrum exhibits two distinct maxima at 303 and 280 nm and a minimum at 262 nm.

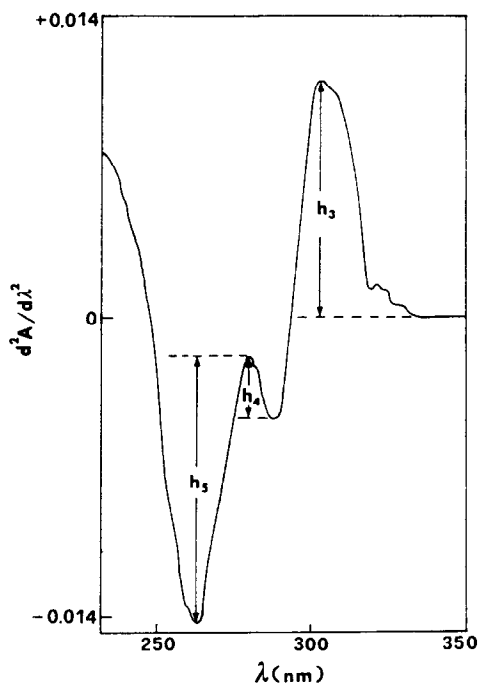
The choice of suitable wavelengths at which to take measurements that are proportional to the cefuroxime and cephalixin concentrations was made after an inspection of the derivative spectra. The measurements made were "peak-to-baseline" and "peak-to-peak" measurements, generally referred to as "graphical measurements" [1].

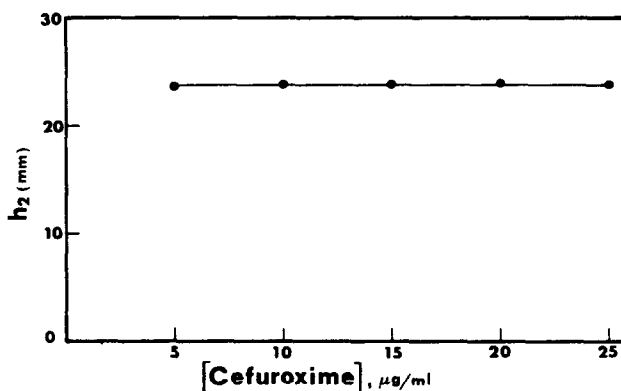
*First derivative mode.* The height of the peak at 296 nm (Fig. 2) is denoted by  $h_1$  (i.e. the distance between the maximum and the baseline), and the distance between the maximum at 276 nm and the intersection of the ordinate through this maximum with the tangent drawn to the left side of the peak at 296 nm is denoted by  $h_2$ . Preliminary experiments showed that  $h_1$  and  $h_2$  were proportional to cefuroxime and cephalixin concentrations, respectively. Moreover, the values of  $h_1$  or  $h_2$  were not affected by the presence of the other cephalosporin. For example, the graph [11] in Fig. 4, shows that

**Figure 2**  
First derivative spectrum of a mixture of cefuroxime sodium ( $15 \mu\text{g ml}^{-1}$ ) and cephalixin sodium ( $20 \mu\text{g ml}^{-1}$ ).



**Figure 3**  
Second derivative spectrum of a mixture of cefuroxime sodium ( $30 \mu\text{g ml}^{-1}$ ) and cephalixin sodium ( $20 \mu\text{g ml}^{-1}$ ).





**Figure 4**

Interaction graph for cephalixin in mixture with cefuroxime ( $4 \mu\text{g ml}^{-1}$ ) by first derivative method ( $h_2$  measurements).

when the cephalixin concentration is constant ( $4 \mu\text{g ml}^{-1}$ ) and the cefuroxime concentration is varied from 5 to  $25 \mu\text{g ml}^{-1}$ , the  $h_2$  value is unaltered.

*Second derivative mode.* The height  $h_3$  at 303 nm in Fig. 3 (peak-to-baseline measurement) was found to be proportional to the concentration of cefuroxime; the heights  $h_4$  at 280 nm and  $h_5$ , distance between the maximum at 280 nm and the minimum at 262 nm (peak-to-peak measurement) were found to be proportional to the concentration of cephalixin. Also, the height  $h_3$  and the heights  $h_4$  and  $h_5$  are independent of cephalixin and cefuroxime concentrations, respectively.

#### *Calibration graphs and statistical analysis of results*

The derivative measurements may be used as easily as ordinary absorbance measurements for the determination of concentration. The linear regression equations calculated for mixtures of cefuroxime sodium and cephalixin sodium in both first and second derivative modes are shown in Table 2, together with the correlation coefficients, the variances and the detection limits, at a level of significance  $P = 0.05$  for 10 standard solutions. The range of concentrations investigated was from 4 to  $30 \mu\text{g ml}^{-1}$  of each cephalosporin and three determinations of each measured value were made.

The ordinate values of the lines of regression ( $H$ ) were calculated from the measurements of  $h$  (mm) read from the chart recordings of the spectra as follows [5-7]:  $H = \text{recorder divisions } (h \text{ mm}) \times \text{scale expansion}/100 \text{ mm full scale}$ .

Beer's Law obtains for concentrations up to 20 and  $28 \mu\text{g ml}^{-1}$  of cephalixin in the first and second derivative modes, respectively, and up to  $30 \mu\text{g ml}^{-1}$  of cefuroxime in both first and second derivative modes.

Tests of significance of the intercept of the lines of regression,  $H = a + bc$ , were performed to establish whether the experimental intercept ( $a$ ) differed significantly from the theoretical value, zero. A simplified method of estimating the differences  $a - 0$  is based on the determination of the quantities  $t = a/s_a$  [20] and their comparison with the corresponding tabular data for  $t$  - distribution:  $s_a$  is defined by the expression:

$$s_a^2 = s_0^2 \Sigma c^2 / n \Sigma c^2 - (\Sigma c)^2$$

**Table 2**  
Statistical analysis of the determination of cefuroxime and cephalixin in mixtures by 1st and 2nd derivative spectrophotometry

Antibiotic	$\lambda$ (nm)	Regression equation	Correlation coefficient	Variance ( $s_0^2$ )	Detection limit ( $\mu\text{g ml}^{-1}$ )
Cefuroxime	296	$H_1 = 9.70\text{E-}04 + 6.78\text{E-}03c$	0.9999	1.02E-06	0.32
Cephalixin	276	$H_2 = 3.20\text{E-}04 + 2.31\text{E-}03c$	0.9996	1.36E-07	0.34
Cefuroxime	303	$H_3 = -1.21\text{E-}04 + 3.61\text{E-}04c$	0.9991	1.99E-08	0.84
Cephalixin	280	$H_4 = 1.69\text{E-}05 + 1.61\text{E-}04c$	0.9987	3.88E-09	0.84
Cephalixin	280/262	$H_5 = -2.01\text{E-}04 + 6.32\text{E-}04c$	0.9991	4.05E-08	0.69

$c$ : concentration of the drug,  $\mu\text{g ml}^{-1}$ .

Number of standard specimens,  $n = 10$ ; level of significance,  $P = 0.05$ .

**Table 3**  
Estimate of significance of the intercept in the calibration data\*

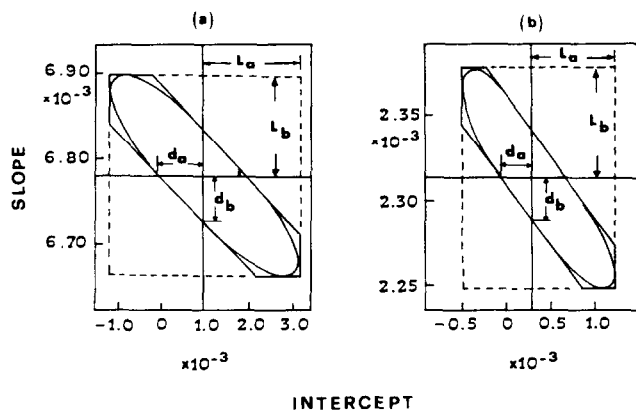
Measurement	Antibiotic	( $a/s_a$ )	$t_p^\dagger$
1st deriv. ( $h_1$ )	Cefuroxime	1.33	2.31
1st deriv. ( $h_2$ )	Cephalixin	1.06	2.31
2nd deriv. ( $h_3$ )	Cefuroxime	1.19	2.31
2nd deriv. ( $h_4$ )	Cephalixin	0.34	2.31
2nd deriv. ( $h_5$ )	Cephalixin	1.24	2.31

\*  $a$  = experimental value of intercept of lines of regression.

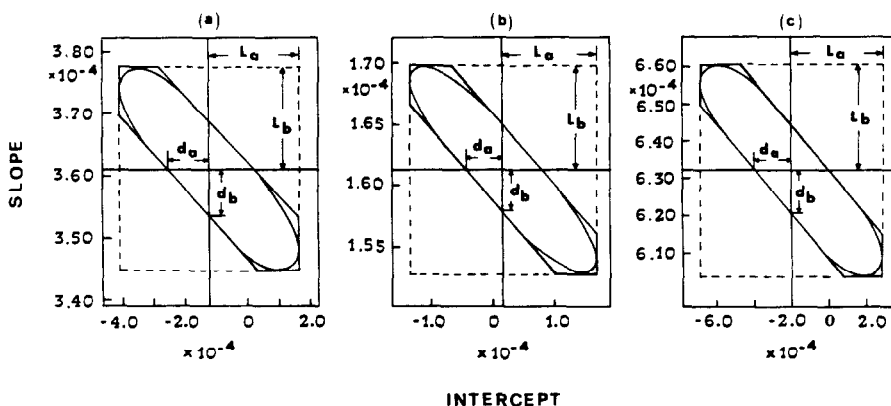
† Theoretical value of  $t$  at  $P = 0.05$  level of significance; no. of degrees of freedom,  $f = 8$ .

in which  $s_0^2$  = variance and  $c$  = concentration of the samples. The results of these calculations are reported in Table 3. The values calculated for  $t$  do not exceed the 95% criterion,  $t_p = 2.31$  ( $t_p$  is Student's coefficient at the selected level of significance), which indicates that the intercepts of all lines of regression are not significantly different from zero and, hence, the proposed methods for determining cefuroxime and cephalixin in mixture are free from errors due to the presence of the other component. However, it should be noted that tests of significance on the estimated slopes and intercepts, when carried out independently of each other, ignore the strong correlation that exists between them. Hence, a more rigorous approach, which requires the construction of a joint confidence region for the possible values of the two parameters, has the further advantage of allowing the evaluation of the calibration graph as a whole.

The 95% joint confidence regions for slopes and intercepts of the regression equations reported in Table 2 are shown in Fig. 5 (first derivative method) and Fig. 6 (second derivative method), constructed by the method of Mandel and Linning [20–21]. These confidence regions are bounded by an ellipse which has the point of best fit as centre (i.e. coordinates  $a$  and  $b$ ). The boundary is dependent on the magnitude of the experimental errors and on the level of significance (0.05, in the present instance) of the probability that the real point lies in the interior of the ellipse.



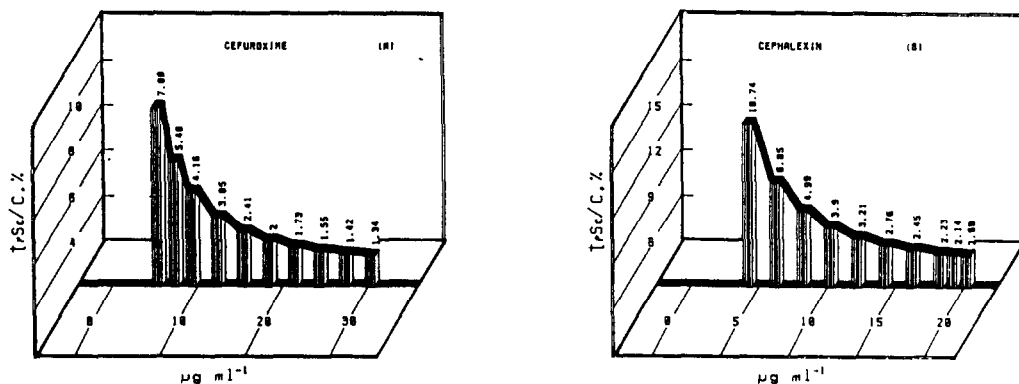
**Figure 5** Joint confidence regions for slope and intercept of regression equations, by first derivative method. (a) cefuroxime sodium ( $h_1$  measurements); (b) cephalixin sodium ( $h_2$  measurements); level of significance  $P = 0.05$ ; number of samples,  $n = 10$ .



**Figure 6** Joint confidence regions for slope and intercept of regression equations, by second derivative method. (a) Cefuroxime sodium ( $h_3$  measurements); (b) Cephalixin sodium ( $h_4$  measurements); (c) Cephalixin sodium ( $h_5$  measurements); level of significance,  $P = 0.05$ ; number of samples,  $n = 10$ .

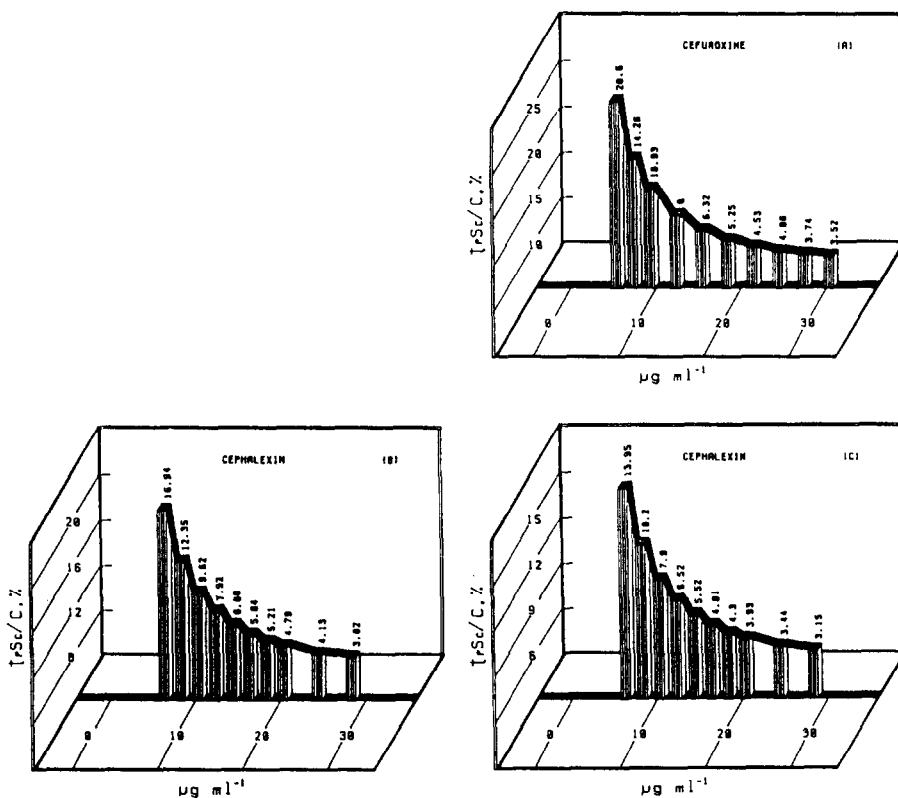
The confidence ellipses were used to confirm the hypothesis  $a = 0$  in the determination of the mixtures of cefuroxime and cephalixin considered above. The question is answered by determining whether the ellipses in Figs 5 and 6 contain points for which the intercept is zero. Such points are on a vertical line through abscissa = 0 and it is evident that they fall well inside the ellipses, thus confirming the hypothesis that  $a = 0$ .

The histograms in Fig. 7 (1st derivative method) and Fig. 8 (2nd derivative method) show the confidence limits [20] for the determination of cefuroxime and cephalixin in a mixture, at  $P = 0.05$  level of significance. These are plotted from calibration data in a particular way [6-8, 15-17, 22, 23], i.e. as uncertainty (%) on concentration (relative error) ( $t_p s_c / c, \%$ ) against the concentration of cefuroxime or cephalixin, respectively. Hence they are a guide to the level of precision that may be expected in the full range of concentrations tested.



**Figure 7**

Histograms of the variation of confidence limits at  $P = 0.05$  level of significance, in the form of uncertainty per cent on the concentration, by first derivative method. (a) cefuroxime sodium ( $h_1$  measurements); (b) cephalixin sodium ( $h_2$  measurements).



**Figure 8**

Histograms of the variation of confidence limits at  $P = 0.05$  level of significance, in the form of uncertainty per cent on the concentration, by second derivative method. (a) Cefuroxime sodium ( $h_3$  measurements); (b) Cephalixin sodium ( $h_1$  measurements); (c) Cephalixin sodium ( $h_3$  measurements).



**Table 4**  
 Replicate determinations on synthetic mixtures of cefuroxime and cephalixin

Measurement	Nominal value ( $\mu\text{g ml}^{-1}$ )		Mean value* ( $\mu\text{g ml}^{-1}$ )		Standard deviation		Relative standard deviation (%)	
	Cefuroxime	Cephalixin	Cefuroxime	Cephalixin	Cefuroxime	Cephalixin	Cefuroxime	Cephalixin
1st deriv. ( $h_1$ and $h_2$ )	18.0	12.0	18.12	11.99	0.0158	0.0016	0.087	0.013
2nd deriv. ( $h_3$ and $h_4$ )	9.0	18.0	8.91	17.86	0.0223	0.0270	0.250	0.151
2nd deriv. ( $h_3$ and $h_5$ )	12.0	16.0	11.96	16.14	0.0286	0.0230	0.239	0.142

\* Mean of five determinations.

To test accuracy and precision of all methods proposed, five successive determinations on synthetic mixtures of cefuroxime and cephalixin were carried out. The results reported in Table 4 show that the accuracy and precision were satisfactory.

#### *Recovery of cefuroxime and cephalixin in pharmaceuticals*

As cefuroxime and cephalixin do not occur together in pharmaceutical preparations, the methods were tested with mixtures of two commercial pharmaceuticals. First and second derivative measurements were applied to the determination of film-coated Ceporex tablets (1 g of cephalixin sodium per tablet) and Kesint injectable powder (1 g of cefuroxime sodium per vial).

Extraction of cephalixin from powdered tablets with methanol (as described in a previous paper [15]) eliminates interference from the tablet excipients. Methanolic extraction, rather than aqueous extraction, was used because the poor solubility of some of the excipients in water caused slight opalescence in the samples and less reproducible results. However, injectable cefuroxime is in a pure form without excipients and consequently prior extraction was unnecessary.

Mixtures of the cephalosporins were prepared by dissolving appropriate amount of the compounds in water. The assays were carried out as described under "Procedure". Table 5 shows the results of five replicate determinations on mixtures of Ceporex and Kesint. The results were in good agreement with the stated content of each cephalosporin.

The method is very simple and illustrates the usefulness of derivative spectrophotometric methods for the rapid, precise and sensitive analysis of mixtures of drugs.

**Table 5**  
Results of the assay of cefuroxime and cephalixin in mixtures of pharmaceuticals\*

Measurement	Mean recovery % <sup>†</sup>	
	Cefuroxime	Cephalixin
1st deriv. ( $h_1$ and $h_2$ )	100.25 ± 0.0710	99.90 ± 0.0178
2nd deriv. ( $h_3$ and $h_4$ )	99.94 ± 0.0961	99.46 ± 0.0226
2nd deriv. ( $h_4$ and $h_5$ )	99.93 ± 0.0903	100.63 ± 0.0254

\*Ceporex tablets, 1 g cephalixin Na/tablet; supplier, Glaxo S.p.A. (Italy). Kesint powder for injections, 1 g cefuroxime Na/vial; supplier, Proter S.p.A. (Italy).

<sup>†</sup>Mean of five determinations ± standard deviation; assay as percentage of label claim.

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